EVALUATING A STRATEGY TO ENHANCE BIODIVERSITY OVER A 426-HA PRAIRIE RESTORATION AFTER 10 YEARS



Abstract

Restoring high levels of biological diversity in landscape restoration work is critical to reestablishing many important ecological functions. In large restorations, this often presents a significant challenge in that seed acquisition is often limited by either financial resources or availability, or both, especially for rare species or those producing little seed. To increase diversity across 426 ha of wet to dry prairie in a 1,113 ha restoration in northern Illinois, 89 species were planted in 60 30-m diameter 'nodes of diversity' to enhance a base seeding of 61 matrix species through ongoing dispersal. Twenty eight nodes were planted with 54 wet prairie species (wet species), 26 nodes with 48 mesic prairie species (mesic species), and six nodes with 33 dry prairie species (dry species). Dispersal potential of node species ranged from low (e.g. gravity dispersal) to high (e.g. wind or animal-vector dispersal). After 10 growing seasons, each node was assessed for species establishment and abundance, with occurrence and abundance assessed in consecutive rings outside each node at 5-m increments to determine if dispersal from nodes was a viable means of increasing diversity across the restoration site.

Eighty node species established in at least one node, with 94.0 % of wet, 79.2 % of mesic, and 87.9 % of dry species found. Nodes averaged 13.7 species in wet prairie, 12.8 species in mesic prairie, and 17.3 species in dry prairie. Not all established species successfully dispersed: 43 % of wet, 45% of mesic, and 46 % of dry species established in the first 5-m ring outside the nodes; 23 % of wet species, 27 % of mesic species, and 30 % of dry species established in the second ring, and 8% of wet species, 8 % of mesic species, and 28 % of dry species established in the third ring.

Establishment potential following dispersal was unrelated to dispersal potential, as no differences were found among the three dispersal potential classes in the proportion of established species that dispersed within any of the three habitat type. However, based on casual monitoring, species with higher dispersal potential did establish well beyond the 60-m diameter area surveyed at each node, but at densities too low to effectively sample, or to make diversity nodes a viable strategy to increase diversity *evenly* across the site over a 10-year period. However, the nodes were successful in introducing species where little seed was affordable or available in densities that promoted the establishment and expansion of viable populations. These nodes now serve as a focus for seed collection for ongoing introductions elsewhere.

Introduction

A general consensus has developed that a positive relationship exists between biological diversity and ecological function (Hooper and Vitousek 1997, Zedler et al. 2001, Cardinale 2002, Lambers et al. 2004, Hooper et al. 2005, Balvanera et al. 2006, Tilman et al. 2006, Hillebrand and Matthiessen 2009, Quijas et al. 2010, Hector et al. 2011, Morin et al. 2011, Midgley 2012, Quijas et al. 2012, Turnbull et al. 2012). From a conservation perspective, particularly in regard to the restoration of globally rare landscapes such as tallgrass prairie, restoring high levels of biological diversity is critical to establishing and maintaining ecological function in developing ecosystems (Schwartz et al. 2000, Callaway 2003, Naeem 2006, Duffy 2009, Webster et al. 2010, Biondini et al. 2011, Isbell et al. 2011, Symstad and Jonas 2011, Tilman et al. 2012, Isbell et al. 2013). One of the most difficult challenges restoring plant species diversity in large-scale restoration is to acquire a sufficient quantity of native plant propagules for each species targeted for reintroduction. Many species that once characterized native landscapes may not be available as they are locally extirpated or rare, and/or they produce very little seed (Howe 1994, Polley et al. 2005). Many species that are not particularly rare within their native habitat may still be difficult or expensive to acquire if little of their habitat remains (Packard 1997). In large restorations, seed acquisition is often limited by both financial resources and availability, especially of rare species or those producing little seed (Rowe 2010, Larson et al. 2011).

To increase prairie species diversity across 426 ha being restored as wet-, mesic-, and dry prairie, we planted 60 730 m² diversity 'nodes' to enhance the matrix of species planted in each of the three prairie habitats through species dispersal over a 10 year period. Our goal was to determine:

- 1) Can the introduction of species in diversity nodes be used as an effective strategy to increase species diversity outside the nodes through ongoing dispersal across a restoration site?
- 2) Are species with higher dispersal potential better at establishing beyond the diversity nodes?
- 3) Are some species better than others at both establishing in nodes and dispersing across the site?

Methods

In 2001, restoration efforts began on 1,113 ha of land that had been planted in corn and soybeans since 1909 to a landscape mosaic of lakes, wetlands, savanna, forest, and prairie (the Sue and Wes Dixon Waterfowl Refuge at Hennepin & Hopper Lakes, now a RAMSAR wetland of international importance). Because of finite financial resources and the inherent scarcity of some species, one of the primary challenges was to acquire sufficient seed to establish high levels of plant diversity on the 426 ha being restored. A matrix of 61 species was planted across the three types of prairie, with each type identified on the landscape based on topography, soil characteristics, and projected hydrology. Species were introduced from seed through 'frost seeding', i.e., seed was planted in late autumn once nights were cold enough to produce frost-heaving, which combined with daytime melting naturally worked seed into exposed mineral soil.

As a strategy to enhance prairie species diversity, 60 30-m diameter 'nodes' of diversity (each 730 m²; Figure 1) were planted with 89 additional species: 28 nodes with 50 wet prairie species (wet species), 26 nodes with 48 mesic prairie species (mesic species), and six nodes with 33 dry prairie species (dry species). The number of nodes was proportional to the area of each type of prairie planted. Nodes were located across the landscape objectively to insure that at least one node was found in each contiguous section of prairie habitat, i.e., each discrete habitat unit had at least one node from which node species could potentially disperse (Figure 2). Consequently, node density was related to the spatial distribution of all habitats across the landscape mosaic. Each node was marked with a steel stake and the coordinates recorded with GPS. The quantity of seed per species varied among species in each node based on cost and availability (Table 1), but in all cases was much less than that of the matrix species planted.

The dispersal potential of each node species was subjectively assigned to one of three dispersal classes based on their physical characteristics. Species ranged from low potential (e.g. gravity dispersal with the heavy round seeds of *Baptisia leucantha*) to high potential (e.g. wind dispersal aided by the fluffy pappus on seeds of Asclepias tuberosa, or animal-vector dispersal with the sticky seeds of *Desmodium illinoense*). Some species were categorized as intermediate due to seed characteristics that appear to promote longer dispersal under specific conditions, e.g. the fine seeds of Penstemon digitalis which are shaken from capsules in wind. After 10 growing seasons, each node was assessed for species establishment and abundance, with abundance assessed in consecutive rings outside each node at 5-m increments to determine if dispersal from nodes was a viable means of increasing diversity evenly across the restoration site. Of the 60 nodes planted in 2002, 53 were sampled in 2012: 26 wet, 23 mesic, and four dry (seven were lost, e.g. one suffering an invasion of Canada thistle was eradicated). Differences among species with different dispersal potentials were assessed with a non-parametric Kruskal-Wallis test, with post hoc Bonferroni pairwise comparisons calculated to assess differences among percent-dispersal means.



Figure 1. Layout of 730 m² diversity nodes planted in 2002 (15-m diameter circle, in green), with consecutive rings at 5 m intervals that were sampled to record dispersal outside each node (rings at 15 to 20-, 20 to 25-, and 25 to 30 m from center).



Figure 2. Distribution of 53 30-m diameter diversity nodes surveyed in 426 ha of wetmesic-, and dry prairie in 2002 at the 1,113-ha Sue and Wes Dixon Waterfowl Refuge at Hennepin & Hopper Lakes.

Eighty node species established in at least one node, with 94.0 % of wet, 79.6 % of mesic, and 78.4 % of dry species found. Nodes averaged 13.7 species in wet prairie, 12.8 species in mesic prairie, and 17.3 species in dry prairie. Not all species that established successfully within each node dispersed (Table 2). Fewer numbers of species dispersed into each consecutive ring surveyed outside the nodes, and most species were not found more than 15 m beyond the outer edge of the nodes, indicating that most dispersal after 10 years was relatively short distance. There was a notable exception in the wet prairie habitat where two species (*Desmanthus illinoense* and *Bidens aristosa*) established in high densities within the footprint of a few surface drainage ways that resulted in longer term moist soil conditions. These species have colonized large portions of the site where those specific conditions have been found. One unexpected outcome was the greater number of species established in dry prairie nodes and the greater proportion of dry prairie species that dispersed into each consecutive ring surveyed despite the generally low recruitment observed in dry prairie in moderate to dry years.

Table 2. The mean proportion of established species that dispersed to each consecutive ring surveyed outside the nodes within each habitat.

Some species did not establish anywhere and others that did establish failed to disperse outside the nodes, while some species were quite good at recruitment as more individuals were found outside the nodes than within (Table 3). Some of these species are relatively conservative and most likely encountered in less-disturbed communities, such as *Liatris pycnostachya* or *Echinacea pallida*, indicating that at least some conservative species can be introduced at lower densities and still populate a restoration. This appears to have happened more consistently around wet habitat nodes, perhaps where soil moisture was less of a constraint to recruitment than in the drier areas. This further suggests a subset of species that could be planted from seed in lower densities, while other species that might be better introduced as plugs.

Gary Sullivan¹, William Sluis², Izabella Redlinski¹ ¹ The Wetlands Initiative, Chicago, IL, USA ² Trine University, Angola, IN, USA

seed / node

12,857

8,100

934 1,229

2,000

4,571 16,071

19,643

5.714

3,571 2,250

20,527

1,500 1,571

176

411

6,714 10,045 1,429

17,143 12,857

357

625 83,571

6,429

23,821 134

10,714

3,143

7,429 800

651 493

Table 1. Identity and quantity of seed for each species planted in a wet-, mesic, or dry prairie diversity node. Note that some species appear in more than one

Wet	prairie	node

wet species	seed / node	mesic species	seed / node	dry species	
galinus purpurea	10,714	Allium cernuum	3,691	Allium stellatum	
Illium cernuum	3,257	Asclepias sullivantii	83	Antennaria plantaginifolia	
sclepias incarnata	86	Baptisia leucantha	911	Asclepias hirtella	
sclepias sullivantii	115	Baptisia leucophaea	120	Asclepias tuberosa	
ster praealtus	1,021	Bouteloua curtipendula	5,357	Aster sericeus	
Baptisia leucantha	911	Bromus kalmii	9,821	Astragalus canadensis	
Bidens aristosa	321	Carex bicknellii	300	Bouteloua curtipendula	
Bidens coronata	446	Cassia marilandica	971	Bromus kalmii	
Boltonia asteroides	25,313	Coreopsis tripteris	1,250	Carex bicknellii	
Cacalia plantaginea	54	Dalea candida	10,179	Carex muhlenbergii	
Carex buxbaumii	189	Dalea purpurea	2,571	Coreopsis lanceolata	
Cicuta maculata	321	Desmanthus illinoensis	750	Coreopsis palmata	
Coreopsis tripteris	1,250	Desmodium canadense	381	Coreopsis tripteris	
Desmanthus illinoensis	750	Desmodium Illinoense	768	Dalea candida	
Desmodium canadense	454	Dodecatheon meadia	1,071	Desmanthus illinoensis	
leocharis erythropoda	583	Echinacea pallida	3,714	Desmodium canadense	
leocharis obtusa	357	Euphorbia corollata	429	Desmodium illinoensis	
Sentiana andrewsii	2,690	Gentiana andrewsii	2,640	Echinacea pallida	
lelenium autumale	4,643	Gentiana flavida	5,000	Euphorbia corollata	
lypericum pyramidatum	33,750	Gentiana puberulenta	6,214	Helianthus rigidus	
ris virginicus	89	Helianthus rigidus	205	Hypericum sphaerocarpo	
uncus dudleyi	8,929	Heuchera richardsonii	2,500	Koeleria cristata	
uncus effusus	35,714	Hypericum pyramidatum	33,750	Lespedeza capitata	
uncus torreyi	2,857	Hypericum sphaerocarpon	3,357	Liatris aspera	
iatris pycnostachya	1,286	Koeleria cristata	5,022	Monarda punctata	
obelia spicata	1,527	Liatris aspera	10,000	Panicum leibergii	
ycopus americanus	3,872	Liatris pycnostachya	1,286	Parthenium integrifolium	
ycopus uniflorus	30,179	Parthenium integrifolium	750	Penstemon digitalis	
ysimachia quadriflora	4,259	Pedicularis canadensis	943	Penstemon pallidus	
ythrum alatum	18,214	Physostegia virginiana	209	Potentilla arguta	
Aentha arvensis	10,714	Potentilla arguta	12,321	Rosa carolina	
/imulus ringens	44,768	Pycnanthemum pilosum	3,304	Solidago nemoralis	
arthenium integrifolium	500	Pycnanthemum virginianum	92,714	I radescantia ohiensis	
hysostegia virginiana	273	Rosa carolina	134		
Pycnanthemum virginianum	91,143	Silphium integrifolium	386		
Rosa carolina	134	Silphium perfoliatum	150		
Rudbeckia fulgida sullivantii	3,654	Silphium laciniatum	1,850		
Rudbeckia triloba	5,009	Silphium terebinthinaceum	1,000		
Scirpus atrovirens	348,286	Sisyrinchium albidum	1,367		
Scirpus pendulus	8,957	Solidago graminifolia	2,500		
Scirpus validus	1,107	Solidago juncea	7,857		
Senecio pauperculus	1,250	Solidago nemoralis	5,357		
Silphium integrifolium	386	Solidago riddellii	9,964		
Silphium laciniatum	1,084	Solidago speciosa	1,357		
Silphium perfoliatum	150	Tradescantia ohiensis	857		
Silphium terebinthinaceum	1 000	Vernonia fasciculata	4 371		
Solidado graminifolia	2 500	Veropicastrum virginicum	57 286		
	9 96/	Zizea aurea	1 768		
	6 420		1,700		
	1.004				
	1,904				
	/14				
remonia fasciculata	5,229				
eronicastrum virginicum	57,286				
izea aurea	1 768				

Mesic prairie node
10 - 1
Dry prairie node being surveyed



Results and Discussion

habitat	15-20m	20-25m	25-30m
wet	0.430	0.225	0.078
mesic	0.449	0.270	0.081
dry	0.457	0.301	0.278

There were significant differences among the three dispersal potential classes in the proportion of species planted that established across all habitat types (Figure 3). A significantly smaller proportion of species established that were ranked in the highest dispersal potential class, with the greatest proportion of species establishing within the nodes from the group ranked as intermediate in dispersal potential (intermediate > low > high). However, there were no differences among the three dispersal potential classes in the proportion of planted species that dispersed outside the nodes, suggesting that dispersal potential (as defined in this experiment) is not a good predictor of successful establishment following dispersal. Since successful dispersal (establishment following dispersal) was not related to dispersal mechanism, the choice of species for introduction at lower densities (to disperse and increase from recruitment) should be independent of dispersal mechanism, and based on the characteristics of individual species and how they interact with a given site.

Figure 3. Box plot of the proportion of each species planted that established from each dispersal potential class across all habitat types (A), with a box plot of the proportion of species planted that dispersed outside the nodes among the three dispersal potential classes (B). Overall differences among means assessed with a Kruskal-Wallis analysis, with differences among means assessed with post hoc Bonferroni pairwise comparisons



No differences were found among the three dispersal potential classes in the proportion of established species that dispersed within any of the three habitat types (Table 4, Figure 4). Although the mean proportion of species that dispersed from each dispersal potential class differed among nodes, success varied widely among nodes and appeared to be strongly influenced by local conditions unrelated to species or group identity. Not all wet prairie nodes were equally wet, nor dry prairie nodes equally dry. Soil characteristics varied among nodes as well (e.g. sandy vs. clayey), leading to differences in the conditions influencing successful establishment. Likely of equal importance was local community dynamics, which varied widely in both invasive pressure, and the relative densities of and among established species. This again suggests that dispersal mechanism is not a good predictor of successful establishment following dispersal locally, despite some species having a greater potential for long distance dispersal.

Table 3. Mean ratio of dispersed to established individuals (D = number dispersed outside the node / number established within the node) for each wet-, mesic-, and dry prairie species. No value indicates that no individuals established, while 0.000 means that no individuals dispersed outside the node.

wet species	D	mesic species	D	dry species	D
	4 868	Desmodium canadense	8 603	Penstemon digitalis	1 820
Solidado graminifolia	4.000	Liatris pycnostachya	1 208	Boutelous curtipendula	1 364
Liatris pycnostachya	2 269	Desmanthus illinoensis	0.942	Desmodium illinoensis	1.004
Silphium perfoliatum	1 776	Vernonia fasciculata	0.882	Echinacea pallida	1.071
Silphium integrifolium	1.387	Desmodium Illinoense	0.820	Carex mublenbergii	0.917
Desmanthus illinoensis	1.329	Coreopsis tripteris	0.669	Monarda punctata	0.833
Bidens aristosa	1 243	Asclepias sullivantii	0.647	Koeleria cristata	0.617
Mentha arvensis	0.833	Dalea purpurea	0.630	Penstemon pallidus	0.563
Lycopus americanus	0.798	Solidago nemoralis	0.623	Coreopsis lanceolata	0.561
Scirpus pendulus	0.595	Zizea aurea	0.583	L espedeza capitata	0.442
Scirpus validus	0.577	Echinacea pallida	0.361	Coreopsis palmata	0.321
Silphium terebinthinaceum	0.506	Bromus kalmii	0.333	Desmodium canadense	0.188
Silphium laciniatum	0.302	Veronicastrum virginicum	0.288	Coreopsis tripteris	0.167
Physostegia virginiana	0.256	Silphium perfoliatum	0.229	Parthenium integrifolium	0.167
Asclepias incarnata	0.235	Parthenium integrifolium	0.217	Asclepias tuberosa	0.143
Zizea aurea	0.223	Hypericum pyramidatum	0.167	Allium stellatum	0.091
Cicuta maculata	0.212	Silphium laciniatum	0.147	Potentilla arguta	0.042
Eleocharis ervthropoda	0.188	Koeleria cristata	0.127	Solidago nemoralis	0.014
Vernonia fasciculata	0.160	Silphium terebinthinaceum	0.103	Dalea candidum	0.012
Helenium autumale	0.152	Baptisia leucantha	0.093	Antennaria plantaginifolia	0.000
	0.125	Pycpanthemum virginianum	0.083	Asclenias hirtella	0.000
Tradoscantia obionsis	0.125	Solidago spociosa	0.000	Astor sorioous	0.000
Roltonia astoroidos	0.123	Silobium integrifelium	0.009	Astragalus canadonsis	0.000
Coreopsis tripteris	0.114	Tradescantia obiensis	0.007		0.000
Scirpus atrovirons	0.100	Routoloua cuttinondula	0.003		0.000
Bantisia leucantha	0.103	Bycpanthemum pilosum	0.056		0.000
Pychanthemum virginianum	0.070	Carex bicknellii	0.050	Liatris aspera	0.000
Veronicastrum virginiarium	0.070	Bantisia leucophaea	0.000	Rosa carolina	0.000
Mimulus ringens	0.043	Dalea candidum	0.000	Tradescantia obiensis	0.000
	0.042	Euphorbia corollata	0.000	Bromus kalmii	0.000
	0.000	Gentiana flavida	0.000	Desmanthus illinoensis	
Allium cernuum	0.000	Gentiana puberulenta	0.000	Hypericum sphaerocarpon	
Asclenias sullivantii	0.000	Hypericum sphaerocarpon	0.000	Panicum leibergii	
	0.000	Physicstogia virginiana	0.000	T anicum leibergi	
	0.000	Physiostegia virginiaria Rosa carolina	0.000		
	0.000	Solidago graminifolia	0.000		
Gentiana andrewsii	0.000	Solidago juncea	0.000		
Hypericum pyramidatum	0.000		0.000		
Iris virginicus	0.000	Cassia marilandica			
Juncus dudlevi	0.000	Dodecatheon meadia			
Lobelia spicata	0.000	Gentiana andrewsii			
l vsimachia quadriflora	0.000	Helianthus rigidus			
Parthenium integrifolium	0.000	Heuchera richardsonii			
Rosa carolina	0.000	Liatris aspera			
Solidago riddellii	0.000	Pedicularis canadensis			
Teucrium canadense	0.000	Potentilla arguta			
Thalictrum dasvcaroum	0.000	Sisvrinchium albidum			
Aster praealtus		Solidago riddellii			
Bidens coronata					
Eleocharis obtusa					
Lycopus uniflorus					
Rudbeckia fulgida					
Rudbeckia triloba					
Senecio pauperculus					

Figure 4. Box plots of the mean proportion of established species in each node that dispersed outside the node among the three dispersal potential classes in each habitat. In each case, differences among dispersal classes were not significant (Kruskal-Wallis).



Table 4. The mean proportion of established species of each dispersal type that dispersed to each consecutive ring surveyed outside the nodes within each habitat. Differences among dispersal types for each habitat type at each consecutive ring were not significant (Kruskal-Wallis).

dispersal ring	habitat	dispersal type			
		1	2	3	
	wet	0.387	0.384	0.574	
15-20 m	mesic	0.323	0.466	0.479	
	dry	0.454	0.526	0.250	
	wet	0.280	0.174	0.356	
20-25 m	mesic	0.228	0.250	0.375	
	dry	0.342	0.324	0.188	
	wet	0.126	0.050	0.091	
25-30 m	mesic	0.046	0.053	0.158	
	dry	0.292	0.304	0.188	

Conclusions

Dispersal from diversity nodes was not a viable strategy to increase species diversity evenly across the 426-ha site over a 10year period. However, the nodes were successful in introducing species locally where little seed was affordable or available, and these nodes now serve as a focus for seed collection efforts to support ongoing introductions elsewhere. Furthermore, some species did disperse outside the site to begin 'mini' single-species nodes of dispersal, but at densities too low to effectively sample, or to determine origin. Although many species can and will disperse effectively, 10 years is too short a period for this strategy to effectively increase plant diversity evenly across a large restoration site.

In general, species with higher dispersal potential were slightly less better at dispersing effectively within the local area outside the nodes, indicating that dispersal potential is not a reliable predictor of establishment success. Establishment success following dispersal is more likely a function of both seed provisioning and the local physical, chemical, and biological characteristics of the site. However, some species, independent of dispersal characteristics, were clearly better at successful establishment following dispersal than others. Where seed quantities are limited, these species would be good candidates for planting from seed at lower densities to naturally populate a site through successful recruitment. Those species that are poor at establishing as well as dispersal would be better introduced as greenhouse grown plugs, especially where little seed is available or affordable.

References

Balvanera, P., A. B. Pfisterer, N. Buchmann, J. S. He, T. Nakashizuka, D. Raffaelli, and B. Schmid. 2006. Quantifying the evidence for biodiversity effects on ecosystem functioning and services. Ecology Letters Biondini, M.E., J.E. Norland, C.E. Grygiel. 2011. Plant Richness-Biomass Relationships in Restored Northern Great Plains Grasslands (USA). International Journal of Ecology 2011, 1-13. Callaway, J. C., G. Sullivan, and J. B. Zedler. 2003. Species-rich plantings increase biomass and nitrogen accumulation in a wetland restoration experiment. Ecological Applications 13:1626-1639.

Cardinale, B. J., M. A. Palmer, and S. L. Collins, 2002, Species diversity enhances ecosystem functioning through interspecific facilitation. Nature 415:426-429. Duffy, J.E. 2009. Why biodiversity is important to the functioning of real-world ecosystems. Frontiers in Ecology and the Environment 7: 437-444. Hector, A., C. Philipson, P. Saner, J. Chamagne, D. Dzulkifli, M. O'Brien, J. L. Snaddon, P. Ulok, M. Weilenmann, G. Reynolds, H. C. J. Godfray. 2011. The Sabah Biodiversity Experiment: a long-term test of the role of tree diversity in restoring tropical forest structure and functioning. Philosophical Transactions of the Royal Society B: Biological Sciences 366:1582, 3303-3315. Hillebrand, H., B. Matthiessen. 2009. Biodiversity in a complex world: consolidation and progress in functional biodiversity research: Consolidation and progress in BDEF research. Ecology Letters 12:12, 1405.

Hooper, D. U., and P. M. Vitousek, 1997. The effects of plant composition and diversity on ecosystem processes, Science 277:1302-1305. Hooper, D. U., F. S. Chapin, J. J. Ewel, A. Hector, P. Inchausti, S. Lavorel, J. H. Lawton, D. M. Lodge, M. Loreau, S. Naeem, B. Schmid, H. Setälä, A. J. Symstad, J. Vandermeer, and D. A. Wardle. 2005. Effects of biodiversity on ecosystem functioning: A consensus of current knowledge. Ecological Monographs 75:3-35. Howe, H.F. 1994. Managing Species Diversity in Tallgrass Prairie: Assumptions and Implications. Conservation Biology 8: 693-704. Isbell, F., P. B. Reich, D. Tilman, S. E. Hobbie, S. Polasky, S. Binder. 2013. Nutrient enrichment, biodiversity loss, and consequent declines in ecosystem productivity. Proceedings of the National Academy of Sciences 110:29, 11911-11916

Isbell, F., V. Calcagno, A. Hector, J. Connolly, W.S. Harpole, P.B. Reich, M. Scherer-Lorenzen, B. Schmid, D. Tilman, J. van Ruijven, A. Weigelt, B.J. Wilsey, E.S. Zavaleta, M. Loreau. 2011. High plant diversity is needed to maintain ecosystem services. Nature 477, 199-202. Lambers, J. H. R., W. S. Harpole, D. Tilman, J. Knops, and P. B. Reich. 2004. Mechanisms responsible for the positive diversity-productivity relationship in Minnesota grasslands. *Ecology Letters* 7:661-668.

Larson, D.L., J.B. Bright, P. Drobney, J.L. Larson, N. Palaia, P.A. Rabie, S. Vacek, and D. Wells. 2011. Effects of planting method and seed mix richness on the early stages of tallgrass prairie restoration. Biological Conservation 144: 3127-3139. Midgley, G.F. 2012. Biodiversity and Ecosystem Function. Science 335:6065, 174-175.

Morgan, J.P., D.R. Collicutt, and J.D. Thompson. 1995. Restoring Canada's Native Prairies: A Practical Manual. Prairie Habitats. Morin, X., L. Fahse, M. Scherer-Lorenzen, H. Bugmann. 2011. Tree species richness promotes productivity in temperate forests through strong complementarity between species. Ecology Letters, 14: 1211–1219. Naeem, S. 2006. Biodiversity and ecosystem functioning in restored ecosystems: Extracting principles for a synthetic perspective. Pages. 210-237 in D. Falk, M. Palmer, and J. Zedler, editors. Foundations of restoration ecology, Island Press, Washington, DC, USA.

Packard, S., 1997. Interseeding. pp. 163-192. In S. Packard and C.F. Mutel (eds.) The Tallgrass Restoration Handbook for Prairies, Savannas, and Woodlands, Island Press, Washington, D.C. Polley, H. W., J.D. Derner, and B.J. Wilsey. 2005. Patterns of Plant Species Diversity in Remnant and Restored Tallgrass Prairies. Restoration Ecology 13: 480–487 Quijas, S., B. Schmid, P.Balvanera. 2010. Plant diversity enhances provision of ecosystem services: A new synthesis. Basic and Applied Ecology 11:7, 582-593. Quijas, S., L. E. Jackson, M. Maass, B. Schmid, D. Raffaelli, P. Balvanera. 2012. Plant diversity and generation of ecosystem services at the landscape scale: expert knowledge assessment. Journal of Applied Ecology **49**:4, 929-940.

Rowe, H.I. 2010. Tricks of the Trade: Techniques and Opinions from 38 Experts in Tallgrass Prairie Restoration. Restoration Ecology 18: 253–262. Schwartz, M. W., C. A. Brigham, J. D. Hoeksema, K. G. Lyons, M. H. Mills, and P. J. van Mantgem. 2000. Linking biodiversity to ecosystem function: Implications for conservation ecology. Oecologia 122:297-305. Symstad, A.J., J.L. Jonas. 2011. Incorporating Biodiversity Into Rangeland Health: Plant Species Richness and Diversity in Great Plains Grasslands. Rangeland Ecology & Management 64:6, 555-572. Tilman, D., P. B. Reich, and J. M. H. Knops. 2006. Biodiversity and ecosystem stability in a decade-long grassland experiment. Nature 441:629-632. Tilman, D., P. B. Reich, F. Isbell. 2012. Biodiversity impacts ecosystem productivity as much as resources, disturbance, or herbivory. Proceedings of the National Academy of Sciences 109:26, 10394-10397. Turnbull, L.A., J. M. Levine, M. Loreau, A. Hector. 2012. Coexistence, niches and biodiversity effects on ecosystem functioning. Ecology Letters (2013) 16: 116-127. Webster, C.R., D.J. Flaspohler, R.D. Jackson, T.D. Meehan, C. Gratton. 2010. Diversity, productivity and landscape-level effects in North American grasslands managed for biomass production. Biofuels 1:3, 451-461. Zedler, J. B., J. C. Callaway, and G. Sullivan. 2001. Declining biodiversity: Why species matter and how their functions might be restored in Californian tidal marshes. BioScience 51:1005-1017.

